



NO-cGMP PATHWAY FACILITATES ACETYLCHOLINE RELEASE AND BRADYCARDIA DURING VAGAL NERVE STIMULATION

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Introduction

Neuronal nitric oxide synthase (nNOS) has been immunohistochemically located in parasympathetic ganglion around the pacemaker of the heart¹. We have shown that inhibition of nNOS or guanylyl cyclase (GC) reduces the heart rate (HR) response to vagal nerve stimulation (VNS) via a presynaptic pathway in the adult guinea pig *in-vitro*². However, the mechanism by which this occurs is unknown. NO has been implicated in increasing the release of acetylcholine in the rat forebrain³ and the cAMP system has been shown to augment acetylcholine release in isolated atria⁴.

We therefore tested the following hypotheses:

1. Does NO act presynaptically via cGMP to augment the exocytotic release of acetylcholine during nerve stimulation?
2. Is this achieved by inhibition of phosphodiesterase (PDE) 3 to increase cAMP - protein kinase A (PKA) dependent phosphorylation of presynaptic calcium channels?

Methods

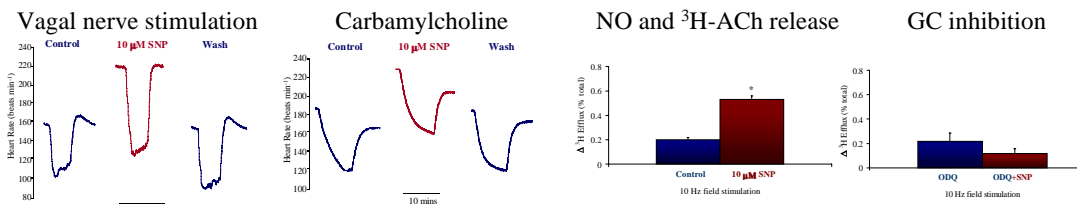
Guinea-pig double atrial/right vagus nerve preparation

The atria and right vagus nerve were dissected from adult (550-750g) female guinea pigs and transferred to a preheated (37±0.2°C) organ bath containing oxygenated Tyrode's solution. Following an equilibration period (60-90 mins), the vagus nerve was stimulated at 1, 3, and 5Hz (10-15V, 1ms pulse duration) for 30 seconds before and after pharmacological interventions.

Measuring right atrial ³H-acetylcholine release

Guinea pig right atrial preparations were placed in a 2ml organ bath at 37±0.5°C and loaded with ³H-choline by repeated (10s every 30s) field stimulation (10Hz, 20V, 1ms pulse duration) for 30mins. Excess radioactivity was washed from the preparation by 30 mins perfusion at 2ml/min. The organ bath Tyrode's solution was then replaced every 3 minutes and its radioactive content measured with a liquid scintillation counter. After 28 mins and again after 43 mins the preparation was stimulated for 1 minute at 10Hz and the change in efflux of radioactivity measured. Experiments were performed in the presence of 50µM hemicholinium 3 to prevent re-uptake of released ³H-acetylcholine and any remaining ³H in the atria was measured at the end of the experiment following incubation with papain (4units/mL). Results are expressed as an efflux of total atrial ³H content.

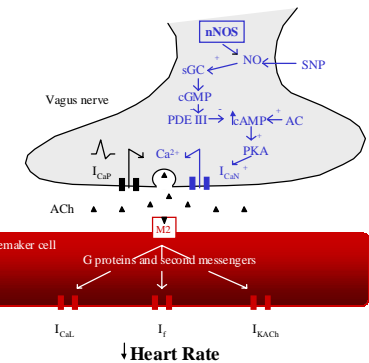
Results 1. NO-cGMP pathway augments acetylcholine release



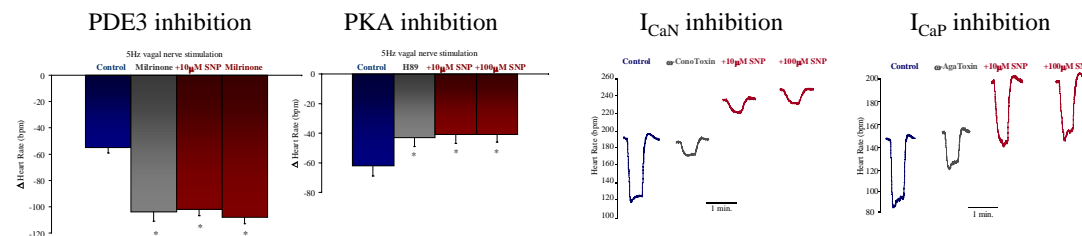
- 10 µM (n=6) or 100 µM (n=7) sodium nitroprusside (SNP) significantly (*p*<0.05) increased the HR response to VNS at 5 Hz (and 3Hz - not shown), but did not increase the HR response to 100 nM carbamylcholine (CCh, n=8). This suggests that SNP acts presynaptically to increase vagal neurotransmission.

- 10 µM (n=4) SNP increased the evoked release of ³H-acetylcholine to field stimulation. However SNP had no effect on the release of acetylcholine (n=4) or the HR response to VNS (n=5) in the presence of the guanylyl cyclase inhibitor ODQ (10 µM).

Summary



Results 2. cGMP-PDE3 pathway increases PKA phosphorylation of I_{CaN}



- SNP had no effect on the HR response to VNS at 5 Hz (n=7) or the release of ³H-acetylcholine (n=4) in the presence of the phosphodiesterase (PDE) 3 inhibitor 1µM milrinone. The protein kinase A (PKA) inhibitor 0.5µM H-89 (n=5), but not the PKG inhibitor 1 µM KT5823 (n=6), abolished the increase in the HR response to VNS with SNP at 5Hz. Similar results were observed at 3Hz VNS - data not shown (**p*<0.05)

- Inhibition of either N-type (with 100 nM ω-Cono Toxin, n=6) or P-type (with 50 nM ω-Aga Toxin, n=5) calcium channels reduced the HR response to VNS at 5 Hz (and 7 and 9 Hz not shown). However SNP did not increase the HR response to VNS in the presence of the N-type calcium channel blocker ω-Cono Toxin.

Conclusions

•SNP releases NO that acts pre-synaptically via a guanylyl cyclase dependent pathway to facilitate vagal release of acetylcholine.

•This is achieved via GMP dependent inhibition of phosphodiesterase 3 to raise levels of cAMP and increase the activity of protein kinase A. Although both N and P-type calcium channels are involved in vagal neurotransmission, protein kinase A phosphorylates N-type calcium channels to facilitate the exocytotic release of acetylcholine.

References

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